

# Detection of Mobile Genetic Elements (MGEs) in Bacterial Genomes

PhD student: Zheng WANG Supervisor: Professor Margaret IP Department of Microbiology, CUHK Date: 3rd Dec, 2013











#### **Mobile genetic elements**



#### **Mobile genetic element**

Any sequence of DNA that is physically moved within an organism genome or between different organisms.

10% - 20% of the Bacterial genome consists of MGEs

#### Horizontal gene transfer

Transfer of genetic material from one organism to another organism that is not its offspring





#### **Mobile genetic elements**



#### **Genomic island**

In a bacterial genome, a cluster of genes for which there is evidence of horizontal origins.

- Prophage
- Integron
- Integrative conjugative element
- Conjugative transposon
- Integrated plasmids





## Importance (X4)

- 1. Frequently associated with microbial adaptations that are of medical and environmental (or industrial ) interest; Metal resistance Antimicrobial resistance Secondary Metabolic properties
- 2. Known virulence factors are over-represented in GIs. The selective loss and regain of GIs could provide an additional means to modulate pathogenicity





#### Importance (X4)

- 3. The spontaneous excision of PAIs has been observed in various pathogens; results in distinct pathogenic phenotypes
- 4.Had a substantial impact on bacterial evolution.





## Methodology

The **Bioinformatics Approaches** for predicting MGEs (especially GIs) with genome sequencing data fall into two broad categories:

- Sequence composition
- *SIGI-HMM* (Hidden Markov Model)
- PAI-IDA.
- Centroid.
- Alien\_Hunter.
- PredictBias.
- PHAST

- Comparative genomics.
- IslandPick
- MobilomeFINDER
- Whole genome alignment

In fact, there are also some **wet-lab methods** to detect MGEs. However, here we just focus on the above well -developed bioinformatic methods.



## Methodology

All of the above methods are based on whole genome sequencing data ; Most of the methods are designed base on GIs sequence and structural Features.

- Sporadic distribution only found in some isolates of a given specie; gene phyletic patterns different with host genome;
- Sequence composition bias

oligonucleotides of various lengths ; GC content; (Traditional Methods)

- Large size (>8 kb)
  - Mobility, phage and virulence genes Over-representation of certain classes of genes and unknown function genes
- Neighbouring tRNA genes ; direct repeats





#### **Methods associated with different Features**

| Feature   | Methods for detection  | Benefits and pitfalls when used for GI prediction  |
|---|--|--|
| Sporadic distribution,<br>instability and an ability to<br>excise spontaneously   | Comparative genomics to identify<br>unique (versus shared) genomic<br>regions  | Multiple closely related sequenced genomes are required for comparison   |
| Sequence composition bias   | Various methods  | False-positive results are obtained owing<br>to a bias in highly expressed genes, and<br>false-negative results are obtained owing<br>to the sequence composition being similar to<br>that of the host genome (which is sometimes<br>the result of amelioration)   |
| Size (usually > 8 kb)   | Comparative genomics to identify<br>large insertions or features such<br>as sequence composition bias in a<br>region over a certain length | Large horizontally acquired regions are easier<br>to predict than regions containing a single<br>gene  |
| Adjacent to a tRNA gene   | Detection of full or partial tRNA<br>genes using BLAST or tRNAscan-SE  | Many GIs are not inserted in or near tRNA genes  |
| Flanked by direct repeats   | Use of repeat finders such as REPuter  | Not all GIs are flanked by direct repeats, and<br>the identification of relevant repeats can be<br>difficult owing to their small size   |
| Over-representation of<br>certain classes of genes such<br>as mobility genes, genes<br>encoding virulence factors,<br>phage-related genes and<br>genes encoding proteins of<br>unknown function | Use of existing genome annotations<br>or searching for similarity to<br>functional databases such as COG'<br>or PFAM                       | Can be used as supporting evidence<br>for GI prediction, and can allow further<br>subclassification of GIs into other MGEs such<br>as prophages or integrated plasmids; but<br>some GIs might have lost all mobility genes, or<br>these genes can be missed because they are<br>not identified by the particular search used |

#### **Overview of genomic island prediction programs**

| Program                      | Description  | Accuracy* and limitations   |
|------------------------------|--|---|
| SIGI-HMM                     | Measures the codon adaptation<br>index and removes ribosomal regions   | <ul> <li>Precision: 92%</li> <li>Recall: 33%</li> <li>Accuracy: 86%</li> <li>The most precise and most accurate program, along with IslandPath-DIMOB</li> </ul> |
| PAI-IDA                      | Measures percentage GC content<br>and dinucleotide and codon usage   | <ul> <li>Precision: 68%</li> <li>Recall: 32%</li> <li>Accuracy: 84%</li> </ul>  |
| Centroid                     | Allows various options, but<br>pentamers are the default   | <ul> <li>Precision: 61%</li> <li>Recall: 28%</li> <li>Accuracy: 82%</li> </ul>  |
| Alien_Hunter                 | Uses variable-length k-mers  | <ul> <li>Precision: 38%</li> <li>Recall: 77%</li> <li>Accuracy: 71%</li> <li>The program with the highest recall, but at the expense of precision</li> </ul>    |
| PredictBias                  | Measures percentage GC content<br>and dinucleotide and codon bias, and<br>predicts PAIs using similarity to a<br>database of virulence genes | <ul> <li>Accuracy measurements could<br/>not be calculated, as the entire<br/>dataset was not available for<br/>download</li> </ul>                             |
| IslandPick                   | Automatically 'picks' default<br>comparison genomes for use in<br>whole-genome alignments  | <ul> <li>The highest agreement with a<br/>data set of GIs that have been<br/>reported in the literature</li> <li>Requires related genomes for use</li> </ul>    |
| MobilomeFINDER <sup>42</sup> | Uses tRNA gene locations and<br>whole-genome alignments to identify<br>GIs   | <ul> <li>Limited to only identifying GIs in<br/>tRNA genes</li> <li>Comparison genomes cannot be<br/>automatically selected</li> </ul>                          |



## **Application example 1**

Identification and characterization of  $\phi$ H111-1: A novel myovirus with broad activity against clinical isolates of *Burkholderia cenocepacia*. (Lynch, K. H.,et al,2013)

- Prophage identification (One of the most important GIs) Using the PHAST method (prophage-finding program Phage Search Tool) to identify prophages in the B. cenocepacia strain H111 genome sequence
- Confirmation of the characterization with laboratory experiments





#### **Methods Selection**

Target Genome status: B. cenocepacia strain H111 only have Draft Genome (gaps unclosed)

#### PHAST

This program accepts either raw reas data or contigs data, however, like all the other GI predict programs, to get a better result, complete genome data are recommended.

Input: 71 available H111 contigs.





#### **PHAST procedures**

- Genome-scale ORF prediction/translation (by GLIMMER)
- Protein identification (by BLAST matching ; annotation by homology)
- Phage sequence identification (byBLAST matching to a phagespecific database)
- tRNA identification
- Attachment site recognition ;
- Gene clustering density measurements (using density-based spatial clustering; DBSCAN)
- Evaluates the completeness of the prophage (give a Score)





#### **PHAST Results**

- GC\_PERCENTAGE; COMPLETENESS: (intact or incomplete, according to SCORE); REGION\_LENGTH and POSITION; CDS;
- In this case, This program identified potential intact prophages (Score >120; total score 150) in contig 43;
- GC content 62% (lower than the H111 GC content of 67%) ;





#### Results



Map of the  $\phi$ H111-1 prophage; the position in the C43 and the CDS;

No putative toxin genes were identified.

#### **Confirmation with laboratory experiments**

- •Transmission electron microscope analysis
- Phage isolation and analysis
- shotgun cloning;

(Lynch, K. H., et al, 2013)





## **Application example 2**

Insight into the specific virulence related genes and toxinantitoxin virulent pathogenicity islands in swine streptococcosis pathogen *Streptococcus equi ssp. zooepidemicus* strain ATCC35246

(Ma, Z. et al, 2013)

 Identification of GIs by Comparative genomics and Sequence composition related methods





**Target strain:** *S. zooepidemicus* strain ATCC35246

NGS: Complete Genome ; 454 Platform.

## **Comparative Genomics**

- 3 Reference genomes : *S. zooepidemicus* MGCS10565 and H70 *S. equi* 4047. (All Complete Genomes)
- identify clusters of genes in target genome that are not present (or scattered )in closely related other 3 Reference genomes
- identify important mobility genes, such as integrases, transposases were present at the boundaries of the region
- GC content (different with the average of whole genome)





#### **Confirm with IslandViewer**

An genomic island predictor that integrates 3methods:

IslandPick,

IslandPath-DIMOB,

SIGI-HMM

GIs which identified by at least 2 methods were marked.





Total 4 GIs associated with pathogenicity and virulence were confirmed

(Ma, Z. et al, 2013)



## **Future Improvements**

• Difficulties :How to Handle un-assembled millions of raw reads .

An increasing proportion of microbial genome sequences are the result of unfinished/unclosed genome sequences Shorter reads might not provide enough signals for sequence composition.

 Trends :The integration of the strengths of previously developed methods coupled with increased genomic database of bacteria and phages.





# References

- 1. Dhillon, B. K., Chiu, T. A., Laird, M. R., Langille, M. G. & Brinkman, F. S. IslandViewer update: Improved genomic island discovery and visualization. *Nucleic Acids Res.* 41, W129-32 (2013).
- 2. Lynch, K. H., Liang, Y., Eberl, L., Wishart, D. S. & Dennis, J. J. Identification and characterization of varphiH111-1: A novel myovirus with broad activity against clinical isolates of. *Bacteriophage* 3, e26649 (2013).
- 3. Ma, Z. *et al.* Insight into the specific virulence related genes and toxin-antitoxin virulent pathogenicity islands in swine streptococcosis pathogen Streptococcus equi ssp. zooepidemicus strain ATCC35246. *BMC Genomics* 14, 377-2164-14-377 (2013).
- 4. Zhou, Y., Liang, Y., Lynch, K. H., Dennis, J. J. & Wishart, D. S. PHAST: a fast phage search tool. *Nucleic Acids Res.* 39, W347-52 (2011).
- 5. Langille, M. G., Hsiao, W. W. & Brinkman, F. S. Detecting genomic islands using bioinformatics approaches. *Nat. Rev. Microbiol.* 8, 373-382 (2010).
- 6. Boyd, E. F., Almagro-Moreno, S. & Parent, M. A. Genomic islands are dynamic, ancient integrative elements in bacterial evolution. *Trends Microbiol.* 17, 47–53 (2009).
- 7. Winstanley, C. *et al.* Newly introduced genomic prophage islands are critical determinants of *in vivo* competitiveness in the Liverpool Epidemic Strain of *Pseudomonas aeruginosa*. *Genome Res.* 19, 12–23 (2008).
- 8. Langille, M. G. & Brinkman, F. S. IslandViewer: an integrated interface for computational identification and visualization of genomic islands. *Bioinformatics* 25, 664–665 (2009).
- 9.Chen, J. & Novick, R. P. Phage-mediated intergeneric transfer of toxin genes. *Science* 323, 139– 141 (2009).





# Thank You !



